



JYOTI NIVAS COLLEGE AUTONOMOUS BANGALORE – 560 095
DEPARTMENT OF BIOTECHNOLOGY
B.Sc. II SEMESTER BIOTECHNOLOGY PAPER II SYLLABUS (2024 SEP BATCH)
MICROBIOLOGY

Course title	MICROBIOLOGY
Course code	24IIBT2T
Course credits	03
Total contact hours	56 Hours
Duration of ESA	03 Hours
Continuous Internal assessment (CIA)	20 Marks
End Semester Examination	80 Marks

COURSE OUTCOMES :

- Compare and contrast about the microbial diversity along with the basic principles of microscopy.
- Illustrate the control of microbial growth by physical and chemical methods and the use of antibiotics and their efficacy testing are emphasized.

LEARNING OUTCOMES:

- Summarize the different parts and working mechanisms of basic light microscope up to electron microscopes with deep knowledge on the sample preparations and staining techniques .
- Demonstrate the sterilization and microbial techniques routinely used in microbial techniques.
- Explain the significance of microbes in understanding antimicrobial agents and its action against various microbes like bacteria, fungi and viral strains.

Unit 1: Microbial diversity & Microscopic techniques

14HRS

Viruses: Structure, classification and reproduction of viruses TMV, adenovirus , bacteriophage (T4 and Lambda) .

Structure and general characteristics of Bacteria: Classification of Bacteria based on Morphology Staining, Nutrition, Environment, biochemical, serology, nucleic acid method. Overview of Bergy manual. Ultra-structure of Bacterial cell. Salient features of Cyanobacteria. **Fungi** -General Structure, classification

Microscopy: Principles of Microscopy resolving power, numerical aperture, working principle and applications of Compound microscope, Dark field microscope, Phase contrast microscope, Fluorescence Microscope, confocal microscope, Electron Microscopes-TEM and SEM.

Unit-2 Microbiological methods

14 HRS

Culture Media: Components of media, natural and synthetic media, chemically defined media, complex media, selective, differential, indicator, enriched and enrichment media **Pure culture methods:** Serial dilution and plating methods (pour, spread, streak); cultivation, maintenance and preservation/stocking of pure cultures; cultivation of anaerobic bacteria **Stains and staining techniques:** Principles of staining. Types of stains-simple stains, structural stains and differential stains(gram staining).

Unit-3 Control of Microorganisms

14 HRS

Definition of terms-sterilization, disinfectant, antiseptic, sanitizer, germicide, microbicidal agents, micro biostatic agent and antimicrobial agent.

Physical methods of control: Principle, construction and applications of moist heat sterilization Boiling, Pasteurization, Fractional sterilization-Tyndallisation and autoclave. Dry heat sterilization-Incineration and hot air oven. Filtration - Diatomaceous earth filter, seitz filter, membrane filter and HEPA; Radiation: Ionizing radiation-y rays and non-ionizing radiation- UV rays.

Chemical methods: Alcohol, aldehydes, phenols, halogen, metallic salts, Quaternary ammonium compounds and sterilizing gases as antimicrobial agents.

Unit-4: Antimicrobial agents

14 HRS

Five modes of action with one example each: Inhibitor of nucleic acid synthesis; Inhibitor of cell wall synthesis; Inhibitor of cell membrane function; Inhibitor of protein synthesis; Inhibitor of metabolism

Antifungal agents: Mechanism of action of Amphotericin B, Griseofulvin

Antiviral agents: Mechanism of action of Amantadine, Acyclovir, Azidothymidine

Antibiotic resistance- MDR, XDR, MRSA, NDM-1

Antibiotic sensitivity testing methods: Disc and Agar well diffusion techniques

Economic and medically important Bacteria and fungi , An Overview on Gut microbiome- prebiotics and probiotics .

BIOTECHNOLOGY PRACTICAL: II
MICROBIOLOGY

Course title	Microbiology
Course code	24IIBT2P
Course credits	02
Total contact hours	3 Hours / week
Duration of ESA	03 Hours
Continuous Internal assessment (CIA)	10 Marks
End Semester Examination	40 Marks

1. To study the principle and applications of important instruments (biological safety cabinets, autoclave, incubator, BOD incubator, hot air oven, light microscope, pH meter) used in the microbiology and Biotechnology laboratory. 1 Unit

2. Sterilization of medium using Autoclave and assessment for sterility. Sterilization of glassware using Hot Air Oven and assessment for sterility 1 Unit

3. Preparation of culture media for bacteria, fungi and their cultivation. 1 Unit

4. Isolation of bacteria and fungi from soil and air 2 Units

5. Plating techniques: Spread plate, pour plate and streak-plate. 1 Unit

6. Colony characteristics study of bacteria isolated from air and soil 2 Units

7. Staining techniques: Bacteria- Gram, Negative staining &
Fungal staining by Lactophenol cotton blue stain 1 Units

8. Study of *Rhizopus*, *Penicillium*, *Aspergillus* using temporary mounts 1 Unit

9. Biochemical Tests - IMVIC, Starch hydrolysis, Catalase test. 2 Units

10. Bacterial cell motility - hanging drop technique. 1 Unit

11) Antibiotic sensitivity test . 1 Unit

Text Books/References:

1. Microbiology fundamentals and applications by Atlas R.M., 2nd Edition, Publisher: McMillan Publishing House, (1998).
2. Microbiology: Principles and Explorations by Black JG and Black LJ. 9th edition. Publisher :John Wiley & Sons (2015).
3. Brock Biology of Microorganisms by Madigan MT, and Martinko JM. 14th edition. Publisher: Pearson education (2014)
4. Microbiology by Pelczar Jr. M.J., Chan E.C.S. & Kreig N.R., 5 th edition , Publisher: Assorted editorial. (2023).
5. Prescott's Microbiology by Joanne Willey, Kathleen sandman & Dorothy wood 12 th edition publisher :Mc Graw- Hill. (2022).